Determination of Arsenic and Cadmium in Sri Lankan Rice Samples by Inductively Coupled Plasma Mass Spectrometry (ICP-MS) Following Microwave Assisted Acid Digestion

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Abstract

Rice is the most important food for the world population including Sri Lanka. However, the toxic metallic elements in rice grains transferred from environment or as an agriculture input often pose great risk to human health because their toxicity, persistence and bioaccumulation in body. Aimed of this study is to investigate the suitability of microwave assisted-acid digestion sample preparation method as a rice sample preparation method as well as to validate the method for monitoring the concentration of arsenic (As) and cadmium (Cd) in rice using Inductively Coupled Plasma Mass Spectroscopy (ICP-MS). The analytical method was validated by measuring several parameters including limit of detection (LOD), limit of quantification (LOQ), linearity, recovery and repeatability. Regarding the sample preparation, recoveries of spiked samples were within the acceptable range from 83.5 to 114.2% for both As and Cd. Validation parameters showed that the method fits for its purpose, being As and Cd within the limit established by CODEX Alimentarius. The method was applied for analyzing 71 rice samples (42 red raw, 27 white raw and 2 parboiled) consumed by Sri Lankan population. Arsenic and cadmium concentrations of all analyzed samples were lower than the established maximum values.

Introduction

Of the ninety (90) naturally occurring elements, seventeen (17) (C, H, O, N, P, S, Ca, Mg, K, Cu, Zn, B, Fe, Mo, Cl, Mn and Ni) are essential for plant life [1], whereas for humans twenty five (25) elements (C, H, N, O, S, P, K, Mg, Ca, Cl, Na, f, Si, V, Cr, Mn, Fe, Co, Ni, Cu, Zn, Se, Mo, Sn and I) are currently known to be essential [2]; in addition, two other elements, Al and Li, have recently been included as possibly essential [3,4]. The twenty five (25) essential elements include four (04) light metals (Na, K, Mg and Ca) and ten (10) heavy metals (Cr, Mn, Fe, Co, Ni, Cu, Zn, Se, Mo and Sn). Food contains most of the above essential metallic elements. These elements are essential in trace quantities for maintenance of cellular processes. The major entry of trace elements into body is through diet. But their excesses, imbalances or deficiencies can results in adverse effects on living system [5,6]. Other metallic elements such as mercury (Hg), lead (Pb), cadmium (Cd), tin (Sn), arsenic (As) etc., have no functional effects in the body and can be harmful to health if foodstuffs containing them are consumed regularly in the diet. Therefore, they are considered as toxic elements [6].

The toxic elements such as cadmium (Cd) and arsenic (As), coming mainly from mining, industrial processes, pesticides, chemical fertilizers and atmospheric decomposition due to the occurrence of industrialization and urbanization without environmental care [7,8]. Toxic elements accumulation in environment is a health hazard due to their persistence, bioaccumulation and toxicity to plant, animal and human beings [9]. An increase in the concentration of a material in a living being over time compared to the concentration of that in the environment is defined as bioaccumulation [8,10]. These toxic elements transported from aquatic media to foodstuffs due to ingestion of foods grown in contaminated soils [11]. It was intensively investigated that uptake of metals by plants depends on the physico-chemical makeup of plant and soil [12]. It has been shown that plants growing in soil contaminated with toxic elements are not capable of preventing their uptake and accumulation in the plant tissue, but are capable of restricting them only [13].

Toxic elements are considered highly hazardous to human health and they may cause acute and chronic poisoning. The principal toxic effect of cadmium (Cd) is its toxicity to the kidney, although it has also been associated with lung damage (including induction of lung tumours) and skeletal changes in occupationally exposed populations. Cadmium is relatively poorly absorbed into the body, but once absorbed is slowly excreted, like other toxic metals and accumulate in the kidney causing renal damage. The kidney of the food animals is a major source of cadmium in the diet although lower levels are found in many foods [6,7]. Arsenic exists both in inorganic and inorganic forms and also in different valence states. Inorganic arsenic is significantly more toxic than organic arsenic compounds such as dimethylarsinate and in turn the trivalent forms of arsenic, e.g. arsenic trichloride, are more toxic than the pentavalent arsenate. The latter are considered to be toxic only after metabolic conversion to the trivalent form of arsenic [6,7]. It has been reported that population are extensively vulnerable to exposure of these toxic elements because of their immature renal system and exhibit a narrow tolerance to these non-essential elements [14]. Therefore some international organizations such as Food and Agricultural Organization (FAO) or World Health Organization (WHO) have identified the Maximum allowable concentration (MAC) in rice [10] and maximum limits in rice [15,16].

Rice (Oryza sativa L.) is the most important food for the world population [17] and one of the most consumed cereals in the world [18], it is part of the staple diet of the world population; it is considered as the most important source of nutrients for billions of people around the world [19]. Rice, being a staple in the diet of many people in Asian countries, supplies a major proportions of the total daily intake of major and trace elements by these people [5] and according to report of FAO, rice provides 30.0% of energy and 20.0% of the daily requirement of protein for adults [10,8,18]. Rice is a rich source of manganese, selenium, magnesium and other essential minerals necessary to body [10]. According to the literature review carried out by Meharg and co-workers, rice roots and grains take up a lot of arsenic than wheat and barley from the soil, even though wheat and barley have been found growing on much more contaminated soil compared to rice [20].

High concentrations of toxic elements are found in rice when compared to other plant grown under the normal conditions. Many toxic elements accumulation in rice is associated with the plant characteristics and its cultivation, as it is usually grown in flooded or very humid areas, which optimize the transfer of such elements from soil to the plant [21]. Only fish and seafood may carry higher concentration of arsenic than rice; however, while arsenic in rice occurs manly as inorganic arsenic species, which are very toxic, arsenic in fish and seafood occurs primarily as organic spices, which are less toxic [21,22].

Various modern developments in agriculture and technology can affect the trace element content of food. Because of this, concerns have increased over the contamination of crop (rice) plants with various chemical elements by introducing of mechanized farming, the application of agro-chemicals and the processing of foodstuffs (rice) during and after harvest. For instance, water used for irrigation during the cultivation together with the trace element contaminated soil. During the postharvest processing, extent of milling of grains is known to affect the trace element content of the cereals such as rice. Milling not only removes the outer layers of the grain, which are rich in minerals but also could potentially contaminate the rice grain with trace elements [23]. Another widely used postharvest practice is parboiling. In this process, the paddy soaked in ambient water for some time. The concern has been raised about possible contamination of paddy by toxic metals during the process of soaking, if clean water is not used. In order to minimize the adverse impact of these practices, it is important to measure and continuously monitor the concentration levels of trace elements in foodstuffs such as rice [23], as it comprises more than 50% of the average daily intake of the people in Sri Lanka. It is also important to note that some people consume rice or rice- based products for all three meals.

The determination of elements in food samples traditionally been performed by digestion with acid or acid mixtures, which is time consuming and involves some potential drawbacks such as analyte losses and sample contaminations due to contaminated environment. The microwave assisted-acid digestion sample preparation method is also an adequate determinative technique, but it demands skilled sample handling, much time to cool the reactor before opening [24]. The main advantage of microwave assisted-acid digestion sample preparation method is its requirement of a small amount of mineral acid and a reduction in the production of nitrous vapours and reduced the risk of the personnel exposure to it [25].

In this context, the scope of this study was to investigate a suitability of microwave assisted-acid digestion sample preparation method as a rice sample preparation method as well as to validate a method for monitoring the concentration of arsenic and cadmium in rice using Inductively Coupled Plasma Mass Spectroscopy (ICP-MS). The proposed method was applied for analysing different type of rice types (red/brown raw rice and white raw rice) coming from various regions of Sri Lanka in order to assist the Sri Lankan regulatory bodies to establish the maximum levels of cadmium and arsenic in rice in Sri Lanka.

Materials and Methods

Reagent and Analytical Solutions

High purity deionized water (resistivity 15 megohmcm⁻¹) that was obtained from a Elix Essential 5 (UV) millipore water purification system (Millipore SAS, Molsheim, France) was used throughout this study. Multi-elements standards solution containing cadmium (¹¹¹Cd) and Arsenic (⁷⁵As) at 10mg/L, multi-element internal standard solution containing germanium (⁷²Ge) and indium (¹¹⁵In) at 10mg/L and ICP-MS instrument optimization solution (Tuning solution) at 1µg/L (¹⁴⁰Ce - celenium, ⁶⁰Co- cobolt, ⁷L - lithiumi, ¹²Mg - magnesium, ²⁰⁵Tl – thalium and ³⁹Y -yttrium) were purchased from Agilent Technologies (Singapore) and traceability estimated for all three solutions with respect to NIST. All of the used reagents were of analytical grade. Trace SELECTTM 69% nitric acid (HNO₃) (Fluka Aanlytical, Sigma-Aldrich, USA) and ACS 30% hydrogen peroxide (H₂O₂) used in the digestion procedures.

External calibration was performed using a six-point standards curve, of which one is reagent blank (2% HNO_3), prepared by serial dilution of multi-element standards solution with 2% (v/v) HNO_3 . Calibration (analytical) curve concentrations ranged from 1.0 to 50.0µg/L (ppb) for both cadmium and arsenic.

A multi-element (germanium for arsenic and indium for cadmium) internal standard solution with $10\mu g/L$ of each element in 2% (v/v) HNO₃ solution was prepared by dilution of 10mg/L. Internal standard solution was added in-line to the analysed solution through a mixing tee, used to blend in the internal standards with the samples after the peristaltic pumping and before the nebulizer.

Purity of the plasma gas (Ar - argon) and collision gas (He - helium) was greater than 99.999%.

All lab ware was soaked in a 10% (v/v) HNO_3 acid solution bath for 24hr and was rinsed with high purity deionized water. Subsequently, all materials were dried under clean-air at ambient temperature.

Instrumentation

The acid digestion of the rice samples were performed using a commercial high-pressure laboratory microwave system (TOP wave microwave for pressure digestion, analytikjena, Analytik Jena AG, Jena) operating at a frequency 50/60Hz with a microwave power 1,450W. The microwave digestion system was equipped with twelve 100mL pressure vessels made of TFM (tetrafluoromethoxy) (PL100). The maximum operating temperature, pressure and sample weight were 230°C (446°F), 40bar (580psi) and 500mg (0.5g) respectively. The minimum fill volume of acid was 7mL.

All the measurements were conducted using single quadruple ICP-MS (Agilent 7700S Series ICP-MS, Agilent Technologies, Singapore). The instrument equipped with a micro-mist glass nebulizer, a cyclonic glass spray chamber and nickel skimmer cone for x-lens. Standard, blank and sample solutions were delivered using an Agilent I-AS auto sampler. A hexapole collision cell (CC) was used for As and Cd determinations. Internal standards (Ge and In) were added in-line to compensate for any effects from acids or instrument drift (non-spectral interference effects). The isobaric interferences were corrected by using correction formulas, which are included in the software of the ICP-MS instrument (Mass Hunter software - full version A 01.02.). The instrument software allowed the rapid switch between standard mode (no gas, cell vented to mass analyzer chamber) to CC mode (Helium gas mode) while continuously aspirating the sample. The ICP-MS operational conditions are summarized in Table 1 [26].

RF power, W	1550
Plasma gas flow rate, L/min	1.0
Auxiliary gas flow rate, L/min	4.3
Integration time, ms	0.1
Number of readings per replicate	3
Isotopes monitored	⁷² Ge [*] , ⁷⁵ As, ¹¹¹ Cd, ¹¹⁵ In [*]

Table 1: Inductively Coupled Plasma Mass Spectroscopy (ICP-MS) operating conditions

*Internal Standards

Mass resolution, mass calibration, sensitivity and stability of the system was checked daily by using a multi-element tuning solution of 1µg/L (1 ppb) of Ce, Co, Li, Mg, Tl and Y. According to the instrument manufacturer sensitivity and mass calibration was monitoring using analytical lines of Li - Lithium-7, Ac - actinium -89 and Tl - Thallium - 205; doubly charged ratio ($^{70}Ce^{2+/140}Ce$) ($\leq 2.5\%$) and oxide ratio (^{156}CeO - cesium oxide/ ^{140}Ce) ($\leq 3.0\%$) were adjusted to low values. Helium mode was used for determination of concentrations of elements in the solutions. Selected analytical mass lines for the analytes to be analyzed in rice were 75 for arsenic and 111 for cadmium [26].

Calculation of the concentration is generally done automatically by the software of the ICP-MS instrument. The content W, as mass fraction, of the element was calculated in mg/kg as followed:

 $W = \{a \ge v \ge F\} / \{m \ge 1000\}$

Where, a is the content (μ g/L) of the element in the test solution, V is the volume (mL) of the digestion solution after being made up to volume, F is the dilution factor of the test solution and m is the mass of the test portion (g).

Samples and Procedure

Rice Sample Preparation Procedure

Sample preparation procedure is a critical point for the success of the analysis [13], being considered an important source of error in method development. In the present study microwave assisted-acid digestion sample preparation method was investigated under two different digestion conditions (two steps and single step) in order to check their suitability for rice decomposition aiming at determining the arsenic and cadmium concentrations by means of ICP-MS.

Eleven organic rice samples (1kg for each) were randomly selected to this study. Those rice samples (5 red raw rice and 6 white raw rice) were bought from several market places located in Homagama city, Colombo district, Sri Lanka and assuming that the production has avoided the use of synthetic fertilizers and pesticides. Results obtained for them were used to validate the method. Seventy one rice samples (42 red raw rice, 27 white raw rice and 2 white parboiled rice) received from different areas in Sri Lanka were used to method application.

At first, the samples were grinded in an analytical mill (National, Singapore); weighed using analytical balance (Shimadzu, Kyota, Japan).

Microwave Assisted-Acid Digestion

For the acid digestion assisted by microwave radiation, the method given by manufacturer was performed with some modifications. In this study, finely homogenized powdered of white and red/brown rice samples (0.5g of each) were weighed and transferred to the dry, clean TFM digestion vessels (PL 100) specific for the microwave oven used. Before digestion, each sample (11 organic samples) was spiked in order to reach a final concentration of $25\mu g/L$ of each arsenic and cadmium using multi-element standard solution (within the calibration curve concentration range) in the final solution (volume completed to 100.0mL by adding deionized water). Nine millilitres of acid mixture (5 HNO₃:1 H2O₂) was added to each vessel and kept for approximately 10 - 15 minutes to react prior to close the lid. The vessels was closed and placed in the rotor and tightened. The loaded rotor was placed in microwave oven. Two different digestion programmes (two steps and one step respectively) were independently run for 11 samples as shown in Table 2. After completion of the digestion and cooling for 30 minutes, the vessel was opened carefully. The digested solution was filtered and transferred into 100mL volumetric flasks and filled with deionized water up to the mark. The non-spiked samples, same standards solution which was used to spike and blank solution (acid mixture) were digested under the same conditions for each digestion programme. Recovery was calculated for each sample.

Demonsterne	TI	Programme 1		D
rarameters	Unit	Step 1	Step 2	Programme 2
Temperature	°C	160	180	180
Pressure	bar	40	40	40
Microwave Power	%	50	70	70
Ramp	min	10	5	10
Time	min	15	15	15

Table 2:	Digestion	programmes
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Method Validation

For the method validation, the sample solution obtained from digestion programme 4 (Table 2) in microwave oven was selected due to the lesser time and temperature consumption. The analytical method validation was performed by considering the limit of detection (LOD), limit of quantification (LOQ), linearity, precision (repeatability). For all of the calculations, Eurachem [27] requirements were considered.

LODs and LOQs were established as three and ten times, respectively, the standard deviation of eight independently digested rice samples (used same rice type which contained lowest arsenic and cadmium content, for all eight analysis). The digested solution was filtered and transferred into 100mL volumetric flasks and filled with deionized water up to the mark. The concentrations were determined using ICP-MS.

Linearity was established separately by preparing the calibration curves of arsenic and cadmium by employing unweighted least-squares liner regression.

Repeatability of the method was evaluated by analysis eight replicates of lowest arsenic and cadmium contained rice sample (same sample used for estimate LODs and LOQs). The repeatability was calculated as relative standard deviation (RSD%).

The amount of matrix present in the test solution to be analyzed may create more or less significant matrix effect compared to pure multi-element standards [26]. Standard addition curves obtained from a rice sample spiked at different concentration levels from 1.0 to 50.0μ g/L were compared with external calibration curves established from multi-element standards in order to evaluate the matrix effect, according to the method described in AOAC 2013.16 2016, [26,28].

Determination of Arsenic and Cadmium in Rice Samples Received from Different Areas of Sri Lanka

To evaluate the analytical method performance for determining the arsenic and cadmium in different rice types consumed by the Sri Lankan population, seventy one rice samples (42 red raw, 27 white raw 2 parboiled white rice samples) from various cities of the country.

Expanded Uncertainty Calculation

Expanded uncertainty estimation (U₂) for the obtained results was calculated according to the criteria described in Eurachem/CITAC [29]. For this evaluation, the following parameters were considered: the overall run to run variation (precision) of the analytical procedure using duplicate test and recovery study using spiked samples of various types of rice. All balances and important volumetric measuring devices are under regular control. Precision and recovery studies take into account the influence of the calibration of the different volumetric measuring devices used during the investigation. Linearity of the response to the relevant trace element (As or Cd) within the given concentration range is established during the validation studies [29].

Expanded uncertainty was determined by multiplying the coverage factor (k) and the combined uncertainty, $u(W): U(W) = k \ge u(W)$, considering the covering factor k = 2, for a confidence level of 95.45%.

Results and Discussion

Sample Preparation Procedure

The well-known polyatomic interference caused by 40 Ar³⁵Cl⁺ was considered in the arsenic determination by using arithmetic correction given in the software in 7700 series Agilent ICP-MS [26]. According to t-test (p = 0.005, n=4) no statistically significant differences between corrected and uncorrected concentration were found. Similar result was found by Mataveil and co-workers [30] for rice flour certified reference material digested with HNO₃ and H₂O₂, reagents also used in the present study. This result can be explained by the fact that only 2% of the total chlorine contents in rice plants are present in the edible part [31].

Table 3 shows the analytes recoveries in the spiked samples submitted to two different microwave digestion programmes for determining arsenic and cadmium in rice. It shows that recoveries depend on the rice type (red raw or white raw) as well as digestion programme. Mean recoveries varies from 83.5 to 114.2% in digestion programme 2, for both types of rice, indicating that the results for both element determined in the sample submitted to the microwave digestion using programme 2 were within the percentage recommended by FDA [32], which comprise the range between 80 and 120%.

Table 3: Percentage recovery of arsenic and cadmium from spiked rice sample submitted to different microwave digestion programmes as indicated in Table 2.

Direction magnetic	Average Recovery (%)			
Digestion programme	Arsenic Cadmium			lmium
	Red (n=5)	White (n=6)	Red (n=5)	White (n=6)
1	91.9	66.3	67.7	101.7
2	114.2	106.2	85.5	83.5

This finding highlights the possibility of using one step digestion programme containing lesser digestion time, temperature and better recovery. It is an advantage in arsenic determination whose volatility is well known.

Mataveil and co-workers [30] also investigated to the suitability of microwave assisted acid digestion method as a sample preparation method for arsenic, cadmium and lead determination in Brazilian rice. Recoveries ranged from 92.8 to 110.2% and these results are comparable to those found on this investigation. The results found in the present study are in agreement with the data reported by Wei *et al* [33], where acid digestion assisted by microwave radiation was carried out, reaching recoveries of 97.0, 106.0 and 103.0% for arsenic, cadmium and lead respectively.

Method Validation Study

The overall performance of the proposed method for arsenic and cadmium determination in rice is summarized in Table 4.

LODs and LOQs results were considered acceptable as the values were below the established by Code Alimentarious [15,16,30] for arsenic and cadmium for rice and its derivatives: 0.30mg/kg and 0.40mg/kg respectively.

Table 4: LOD, LOQ,	working range and correlation	1 coefficient of calibration	curve,	repeatability for	microwave
	assisted acid digest	ion and ICP-MS method	d.		

Parameters	Unit	As	Cd
LOD	mg/kg	0.004	0.003
LOQ	mg/kg	0.014	0.010
Working range	μg/L	1- 50	1 - 50
Correlation coefficient of the calibration curve (r)		0.9995	0.9996
Repeatability (RSD)	%	4.2	4.7

The linearity of the calibration curve was evaluated by investigation the correlation coefficient (r) of the calibration curve. The r values were higher than the 0.998 [27] for both of the analytes, as shown in Table 4, and the regression analyses showed that the linear correlation between concentration and ratio of instrument responses (cps for calibration standard/cps for internal standard) was significant (p < 0.05).

For both elements, the relative standard deviation, RSD% (repeatability) was compliance with European Commission Decision 2002 [34]. It did not exceed 10.0% as shown in Table 4.

Results were obtained in evaluation of matrix effect are summarized in Table 5.

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Calibratian Course	Recovery (%) - 25 μg/L			
Calibration Curve	Arsenic Cadmium			nium
	Red White		Red	White
External	108.5 - 118.1	98.8 -111.2	82.2 - 88.0	81.8 - 85.0
Standard addition	101.5 - 105.3	97.4 - 111.4	90.5 - 95.5	89.8 - 90.6

As shown in the Table 5, no matrix effect was observed and therefore aqueous standards were used for calibration [26,28].

Therefore, as expected, the use of microwave oven for rice acid digestion was appropriate for determination of the arsenic and cadmium concentration in rice using ICP-MS.

Method Application

The above method was applied for quantifying the concentration of arsenic and cadmium in 71 rice samples (42 red raw, 27 white raw and 2 white parboiled) consumes in Sri Lanka. Results are summarized in Table 6.

Table 6: Concentrations of arsenic and cadmium in the analyzed rice samples [average $(n = 3) \pm U(c)^*$]

Rice Type	As (µg/kg)	Cd (µg/kg)
White raw	146.132 ± 0.438	25.718 ± 0.033
White raw	46.396 ± 0.139	11.748 ± 0.015
White raw	109.247 ± 0.328	$< 10.000^{b}$
White raw	74.881 ± 0.225	14.065 ± 0.018
White raw	-	14.785 ± 0.019
White raw	91.314 ± 0.274	18.512 ± 0.024
White raw	100.885 ± 0.303	10.917 ± 0.014
White raw	116.375 ± 0.349	71.158 ± 0.093
White raw	221.066 ± 0.663	21.921 ± 0.028
White raw	17.185 ± 0.052	89.182 ± 0.116
White raw	96.774 ± 0.290	$< 10.000^{b}$
White raw	80.570 ± 0.242	$< 10.000^{b}$
White raw	42.382 ± 0.127	52.014 ± 0.068
White raw	< 14.000 ^b	31.143 ± 0.040
White raw	44.199 ± 0.133	40.516 ± 0.053
White raw	146.132 ± 0.516	16.316 ± 0.021

White raw	147.420 ± 0.442	31.353 ± 0.041
White raw	$< 14.000^{b}$	$< 10.000^{\rm b}$
White raw	26.428 ± 0.079	21.296 ± 0.029
White raw	120.809 ± 0.362	32.930 ± 0.043
White raw	25.424 ± 0.076	33.714 ± 0.044
White raw	59.948 ± 0.180	10.108 ± 0.013
White raw	43.061 ± 0.129	$< 10.000^{b}$
White raw	$3\overline{7.022 \pm 0.111}$	$3\overline{6.574 \pm 0.048}$
White raw	49.253 ± 0.148	-
White raw	147.437 ± 0.442	-
White raw	38.779 ± 0.116	$< 10.000^{b}$
Parboiled white	121.622 ± 0.365	11.913 ± 0.015
Parboiled white	51.331 ± 0.154	87.531 ± 0.114
Red raw	68.911 ± 0.207	97.787 ± 0.127
Red raw	63.898 ± 0.192	35.343 ± 0.046
Red raw	73.956 ± 0.222	59.841 ± 0.078
Red raw	19.330 ± 0.058	131.327 ± 0.171
Red raw	81.775 ± 0.245	50.935 ± 0.066
Red raw	$< 4.000^{a}$	-
Red raw Red raw	< 4.000 ^a < 14.000 ^b	
Red raw Red raw Red raw	$\frac{< 4.000^{a}}{< 14.000^{b}}$ 18.294 ± 0.055	- - 156.492 ± 0.203
Red rawRed rawRed rawRed raw	$< 4.000^{a} \\ < 14.000^{b} \\ 18.294 \pm 0.055 \\ 29.341 \pm 0.088$	$- \\ - \\ 156.492 \pm 0.203 \\ 19.760 \pm 0.026$
Red rawRed rawRed rawRed rawRed rawRed raw	$< 4.000^{a}$ $< 14.000^{b}$ 18.294 ± 0.055 29.341 ± 0.088 19.837 ± 0.060	$- \\ - \\ 156.492 \pm 0.203 \\ 19.760 \pm 0.026 \\ 238.246 \pm 0.310 \\ \end{array}$
Red rawRed rawRed rawRed rawRed rawRed rawRed raw	$< 4.000^{a}$ $< 14.000^{b}$ 18.294 ± 0.055 29.341 ± 0.088 19.837 ± 0.060 159.843 ± 0.480	$\begin{array}{c} - \\ - \\ 156.492 \pm 0.203 \\ 19.760 \pm 0.026 \\ 238.246 \pm 0.310 \\ 27.056 \pm 0.035 \end{array}$
Red rawRed rawRed rawRed rawRed rawRed rawRed rawRed rawRed raw	$< 4.000^{a}$ $< 14.000^{b}$ 18.294 ± 0.055 29.341 ± 0.088 19.837 ± 0.060 159.843 ± 0.480 86.093 ± 0.258	$\begin{array}{c} - \\ 156.492 \pm 0.203 \\ 19.760 \pm 0.026 \\ 238.246 \pm 0.310 \\ 27.056 \pm 0.035 \\ 112.583 \pm 0.146 \end{array}$
Red rawRed raw	$< 4.000^{a}$ $< 14.000^{b}$ 18.294 ± 0.055 29.341 ± 0.088 19.837 ± 0.060 159.843 ± 0.480 86.093 ± 0.258 24.732 ± 0.074	$-$ 156.492 ± 0.203 19.760 ± 0.026 238.246 ± 0.310 27.056 ± 0.035 112.583 ± 0.146 253.137 ± 0.329
Red rawRed raw	$< 4.000^{a}$ $< 14.000^{b}$ 18.294 ± 0.055 29.341 ± 0.088 19.837 ± 0.060 159.843 ± 0.480 86.093 ± 0.258 24.732 ± 0.074 61.703 ± 0.185	$-$ 156.492 ± 0.203 19.760 ± 0.026 238.246 ± 0.310 27.056 ± 0.035 112.583 ± 0.146 253.137 ± 0.329 140.772 ± 0.183
Red rawRed raw	$< 4.000^{a}$ $< 14.000^{b}$ 18.294 ± 0.055 29.341 ± 0.088 19.837 ± 0.060 159.843 ± 0.480 86.093 ± 0.258 24.732 ± 0.074 61.703 ± 0.185 58.188 ± 0.175	$-$ 156.492 ± 0.203 19.760 ± 0.026 238.246 ± 0.310 27.056 ± 0.035 112.583 ± 0.146 253.137 ± 0.329 140.772 ± 0.183 88.198 ± 0.115
Red rawRed raw	$< 4.000^{a}$ $< 14.000^{b}$ 18.294 ± 0.055 29.341 ± 0.088 19.837 ± 0.060 159.843 ± 0.480 86.093 ± 0.258 24.732 ± 0.074 61.703 ± 0.185 58.188 ± 0.175 54.725 ± 0.164	$-$ 156.492 ± 0.203 19.760 ± 0.026 238.246 ± 0.310 27.056 ± 0.035 112.583 ± 0.146 253.137 ± 0.329 140.772 ± 0.183 88.198 ± 0.115 68.588 ± 0.089
Red rawRed raw	$< 4.000^{a}$ $< 14.000^{b}$ 18.294 ± 0.055 29.341 ± 0.088 19.837 ± 0.060 159.843 ± 0.480 86.093 ± 0.258 24.732 ± 0.074 61.703 ± 0.185 58.188 ± 0.175 54.725 ± 0.164 52.087 ± 0.156	$-$ 156.492 ± 0.203 19.760 ± 0.026 238.246 ± 0.310 27.056 ± 0.035 112.583 ± 0.146 253.137 ± 0.329 140.772 ± 0.183 88.198 ± 0.115 68.588 ± 0.089 61.865 ± 0.080
Red rawRed raw	$< 4.000^{a}$ $< 14.000^{b}$ 18.294 ± 0.055 29.341 ± 0.088 19.837 ± 0.060 159.843 ± 0.480 86.093 ± 0.258 24.732 ± 0.074 61.703 ± 0.185 58.188 ± 0.175 54.725 ± 0.164 52.087 ± 0.156 33.321 ± 0.100	$-$ 156.492 ± 0.203 19.760 ± 0.026 238.246 ± 0.310 27.056 ± 0.035 112.583 ± 0.146 253.137 ± 0.329 140.772 ± 0.183 88.198 ± 0.115 68.588 ± 0.089 61.865 ± 0.080 107.638 ± 0.140
Red rawRed raw	$< 4.000^a$ $< 14.000^b$ 18.294 ± 0.055 29.341 ± 0.088 19.837 ± 0.060 159.843 ± 0.480 86.093 ± 0.258 24.732 ± 0.074 61.703 ± 0.185 58.188 ± 0.175 54.725 ± 0.164 52.087 ± 0.156 33.321 ± 0.100 79.797 ± 0.239	$-$ 156.492 ± 0.203 19.760 ± 0.026 238.246 ± 0.310 27.056 ± 0.035 112.583 ± 0.146 253.137 ± 0.329 140.772 ± 0.183 88.198 ± 0.115 68.588 ± 0.089 61.865 ± 0.080 107.638 ± 0.140 56.402 ± 0.073
Red rawRed raw	$< 4.000^{a}$ $< 14.000^{b}$ 18.294 ± 0.055 29.341 ± 0.088 19.837 ± 0.060 159.843 ± 0.480 86.093 ± 0.258 24.732 ± 0.074 61.703 ± 0.185 58.188 ± 0.175 54.725 ± 0.164 52.087 ± 0.164 33.321 ± 0.100 79.797 ± 0.239 17.371 ± 0.052	$-$ 156.492 ± 0.203 19.760 ± 0.026 238.246 ± 0.310 27.056 ± 0.035 112.583 ± 0.146 253.137 ± 0.329 140.772 ± 0.183 88.198 ± 0.115 68.588 ± 0.089 61.865 ± 0.080 107.638 ± 0.140 56.402 ± 0.073 91.870 ± 0.119
Red rawRed raw	$< 4.000^{a}$ $< 14.000^{b}$ 18.294 ± 0.055 29.341 ± 0.088 19.837 ± 0.060 159.843 ± 0.480 86.093 ± 0.258 24.732 ± 0.074 61.703 ± 0.185 58.188 ± 0.175 54.725 ± 0.164 52.087 ± 0.156 33.321 ± 0.100 79.797 ± 0.239 17.371 ± 0.052 89.978 ± 0.270	$-$ 156.492 ± 0.203 19.760 ± 0.026 238.246 ± 0.310 27.056 ± 0.035 112.583 ± 0.146 253.137 ± 0.329 140.772 ± 0.183 88.198 ± 0.115 68.588 ± 0.089 61.865 ± 0.080 107.638 ± 0.140 56.402 ± 0.073 91.870 ± 0.119 33.046 ± 0.043
Red rawRed raw	$< 4.000^{a}$ $< 14.000^{b}$ 18.294 ± 0.055 29.341 ± 0.088 19.837 ± 0.060 159.843 ± 0.480 86.093 ± 0.258 24.732 ± 0.074 61.703 ± 0.185 58.188 ± 0.175 54.725 ± 0.164 52.087 ± 0.156 33.321 ± 0.100 79.797 ± 0.239 17.371 ± 0.052 89.978 ± 0.270 75.743 ± 0.227	$-$ 156.492 ± 0.203 19.760 ± 0.026 238.246 ± 0.310 27.056 ± 0.035 112.583 ± 0.146 253.137 ± 0.329 140.772 ± 0.183 88.198 ± 0.115 68.588 ± 0.089 61.865 ± 0.080 107.638 ± 0.140 56.402 ± 0.073 91.870 ± 0.119 33.046 ± 0.043 104.314 ± 0.136

Red raw	36.866 ± 0.111	81.745 ± 0.106
Red raw	54.282 ± 0.163	96.094 ± 0.125
Red raw	89.156 ± 0.267	64.956 ± 0.084
Red raw	46.453 ± 0.139	49.491 ± 0.064
Red raw	65.851 ± 0.198	47.319 ± 0.062
Red raw	122.143 ± 0.366	29.489 ± 0.038
Red raw	59.919 ± 0.180	118.302 ± 0.154
Red raw	49.784 ± 0.149	101.137 ± 0.131
Red raw	85.437 ± 0.256	80.202 ± 0.104
Red raw	28.608 ± 0.086	94.695 ± 0.123
Red raw	61.202 ± 0.184	53.370 ± 0.069
Red raw	86.160 ± 0.258	63.743 ± 0.083
Red raw	47.369 ± 0.142	30.674 ± 0.040
Red raw	51.609 ± 0.155	55.534 ± 0.072
Red raw	50.808 ± 0.152	12.166 ± 0.016
Red raw	52.760 ± 0.158	55.799 ± 0.073
Red raw	53.237.±0.113	$< 10.000^{b}$
Red raw	83.549 ± 0.251	53.821 ± 0.070
Red raw	30.502 ± 0.092	41.966 ± 0.055

^a : LOD; ^b : LOQ; [*] U(c): expanded uncertainty, coverage factor
k = 2, for a confidence level of 95.45%

According to Table 6, out of 71 analyzed samples, 67 showed arsenic concentration higher than the LOQ, ranging from 17.185 to 221.066 μ g/kg (0.017 to 0.221mg/kg)

In one red raw rice sample, arsenic concentration was lower than the LOD. In three samples (one red raw rice and two white raw rice), arsenic concentration was lower than the LOQ and higher than LOD. The average arsenic concentration of parboiled rice was higher than that found in other rice types ($86.476\mu g/kg$), followed by white raw rice ($77.014\mu g/kg$) and red raw rice, which had the lowest arsenic concentration ($57.407\mu g/kg$). In the process of rice parboiling, the paddy is soaked in ambient water for some time, it has a potential to contamination of paddy by trace elements and other contaminants during the soaking, if clean water is not used [5].

In one white raw rice sample, arsenic concentration was 221.066µg/kg (0.221mg/kg). This value also lower than the maximum value established by the CODEX Alimentarius [15], which is 300µg/kg (0.30mg/kg). The samples (red raw rice, white raw rice and parboiled rice) were considered satisfactory for human consumption with respect to international limits for arsenic in rice established by the CODEX Alimentarius [15], but in Sri Lanka, there is no maximum limit for arsenic in rice.

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The arsenic concentrations found in the present study were in agreement with those previously, reported for rice in other countries investigations, as shown in Table 7 [30], and also comparable to others in investigation performed in Sri Lanka [5].

Country	Element concentration (mg/kg)	
	As	Cd
China	0.080	0.037
	0.336	0.312
	-	0.15
Bangladesh	0.32	-
	0.321	0.088
India	0.051	0.019
Iran	-	< 0.015
	0.121	-
	0.065	-
United Kingdom	0.187	-
	0.124 (polished)	-
	0.205 (brown)	-
Spain	0.1695	-
Italy	0.18 - 0.28	0.01 - 0.08
Thailand	0.139 - 0.239	-
Japan	0.032 - 0.239	0.013 - 0.043
Turkey	0.0985	0.314
Greece	0.042 - 0.271	-
Jamaica	0.11 - 0.165	< 0.040 - 0.033

Table 7: Concentrations of arsenic and cadmium in rice consumed by the population of various countries, reported in the literature [30]

With respect to cadmium, concentrations of cadmium of all samples were above the LOD; only seven below the LOQ and all other were above the LOQ. According to the Table 6, Cadmium concentrations higher than LOQ were lies in between 10.108 and 257.409 μ g/kg (0.010 and 0.257mg/kg). In none of the analyzed samples, cadmium concentrations exceeded the maximum value of 400 μ g/kg (0.40mg/kg) allowed by the CODEX Alimentarius [16]. Thus referring cadmium contamination, the red raw rice, white raw rice and parboiled rice were considered as satisfactory for human consumption.

In the view of results of present study, an elevated level of cadmium was found in red raw rice samples (average value 79.276 μ g/kg), whilst the lower level of cadmium is found in both white raw rice (average value 23. 292 μ g/kg) and parboiled rice (average value 49.722 μ g/kg).

The average concentrations of cadmium found in the present study were in agreement with those cited in the literature; and they pointed out the evident of occurrence of the highest cadmium concentration in red raw rice than white rice [30,35].

Cadmium concentrations found in the analyzed samples were similar to those reported in other studies carried out in other countries, as outlined in Table 7 [20].

Overall variation of arsenic and cadmium levels observed in different rice samples from Sri Lanka as well as from other countries can generally be attributed to various factors such as varietal differences, soil composition, geographical location, environmental factors, agricultural chemicals, post-harvest processing water quality, growing conditions and quality of trace element analysis [5,30].

Conclusions

Today, industrialization and development occurs without caring the environment and have led to heavy metal contamination of human food. Dietary intake is the main route of exposure to trace elements such as As and Cd for most of people and have drawn a lot of attention. The microwave assisted acid digestion sample preparation method applied here is found to a convenient, rapid, reproducible, minimum amount of acid used and shorter time required sample preparation method (Mean recoveries varies from 83.5 to 114.2% for both elements). The parameters obtained for validation have demonstrated that the method fits for its purpose, which is to quantify the arsenic and cadmium in rice samples by ICP-MS. This information is important for laboratories that execute rice analysis. Regarding the red raw, white raw and parboiled rice samples analyzed, the concentrations are found for arsenic ranging from 17.185 to 221.066µg/kg (0.017 to 0.221mg/kg) and for cadmium ranging from 10.108 and 257.409µg/kg (0.010 and 0.257mg/kg) were lower than the established limits (0.30mg/kg for arsenic and 0.40mg/kg for cadmium in rice) according to Codex Alimentarious. It appears that rice samples from Sri Lanka were considered satisfactory for human consumption regarding the investigated elements.

In Sri Lankan point of view, there are no maximum limits for trace elements in rice. It is better to established limits with considering geological factors, rainfall pattern, types of chemical use, pattern of chemical usage etc. Also, results of this study can be used as baseline values for future comparisons.

Conflicts of Interests

There is no conflict of interests.

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